

# Transfecting Plasmid DNA into COS-7 Cells Using Lipofectamine<sup>™</sup> LTX Reagent

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#### Introduction

Lipofectamine<sup>™</sup> LTX Reagent is a proprietary, animal-origin free formulation for the transfection of DNA into eukaryotic cells with **low cytotoxicity**. This reference provides a recommended procedure to transfect plasmid DNA into COS-7 Monkey Kidney Fibroblasts (ATCC Cat. No. CRL-1651) using Lipofectamine<sup>™</sup> LTX Reagent (Cat. No. 15338-100).

#### Important Guidelines for Transfection

Follow these important guidelines when transfecting DNA into COS-7 cells using Lipofectamine™ LTX Reagent:

- The addition of antibiotics to media during transfection may result in cell death, and has not been tested for COS-7 cells. If you wish to use antibiotics during transfection, test your conditions thoroughly.
- Maintain the same seeding conditions between experiments. Use low-passage cells; make sure that cells are healthy and greater than 90% viable before transfection.
- Transfection can be performed both in the presence or absence of serum. Test serum-free media for compatibility with Lipofectamine™ LTX Reagent.
- Using PLUS™ Reagent (Cat. No. 11514-015) enhances transfection performance in COS-7 cells.
- We recommend Opti-MEM® I Reduced Serum Medium (Cat. No. 31985-062) to dilute the DNA and Lipofectamine™ LTX Reagent before complexing.
- Visit <a href="www.invitrogen.com/transfection">www.invitrogen.com/transfection</a> or contact Technical Service for other specialized transfection protocols (including cell-type specific advice on use of PLUS™ Reagent and antibiotics, and a protocol for vector-based RNAi).
- Lipofectamine<sup>™</sup> LTX Reagent performs well with vector-based RNAi experiments. For siRNA and Stealth<sup>™</sup> RNAi transfections, we recommend Lipofectamine<sup>™</sup> RNAiMAX (Cat. No. 13778-075). Go to <a href="https://www.invitrogen.com/RNAi">www.invitrogen.com/RNAi</a> or contact Technical Service for more information.

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### **Materials Needed**

Have the following reagents on hand before beginning:

- COS-7 cells maintained in DMEM supplemented with L-glutamine (Cat. No. 11965-084), 0.1 mM MEM Non-Essential Amino Acids Solution (Cat. No. 11140-050), and 10% Fetal Bovine Serum (Cat. No. 26140-079). Grow cells at 37°C with 5% CO<sub>2</sub>.
- Plasmid DNA of interest (100 ng/µl or higher)
- Lipofectamine<sup>™</sup> LTX Reagent (store at +4°C until use), and PLUS<sup>™</sup> Reagent (if desired; store at 4°C)
- Opti-MEM® I Reduced Serum Medium
- Appropriate tissue culture plates and supplies

## **Transfection of COS-7 Cells**

Use this procedure to transfect plasmid DNA into COS-7 cells in a **24-well format** (for other formats, see **Scaling Up or Down Transfections**, below). All amounts and volumes are given on a per well basis.

- 1. The day before transfection, trypsinize and count the cells. Plate  $5 \times 10^4$  cells per well in 0.5 ml of complete growth medium. Cell density should be  $50 \sim 80\%$  confluent on the day of transfection.
- 2. For each well of cells to be transfected, dilute 0.5  $\mu g$  of DNA into 100  $\mu l$  of Opti-MEM® I Reduced Serum Medium without serum.
- 3. If using PLUS<sup>™</sup> Reagent: Mix PLUS<sup>™</sup> Reagent gently before use, then add 0.5 µl PLUS<sup>™</sup> Reagent (a 1:1 ratio to DNA) directly to the diluted DNA. Mix gently and incubate for 5-15 minutes at room temperature.
- 4. For each well of cells, dilute 1.25-2.25 μl of Lipofectamine<sup>™</sup> LTX into the above diluted DNA solution, mix gently and incubate for 25 minutes at room temperature to form DNA-Lipofectamine<sup>™</sup> LTX complexes.
- 5. Remove growth medium from cells and replace with 0.5 ml of complete growth medium. Add 100 µl of the DNA-Lipofectamine™ LTX complexes directly to each well containing cells and mix gently by rocking the plate back and forth.
- 6. Complexes do not have to be removed following transfection. Incubate the cells at 37°C in a CO<sub>2</sub> incubator for 18-24 hours post-transfection before assaying for transgene expression.

# **Scaling Up or Down Transfections**

To transfect COS-7 cells in different tissue culture formats, vary the amounts of Lipofectamine<sup>™</sup> LTX Reagent, DNA, cells, medium and PLUS<sup>™</sup> Reagent used in proportion to the relative surface area, as shown in the table (amounts given on a per well basis).

Culture vessel	Surface area per well <sup>1</sup>	Volume plating medium	Cells per well	Volume dilution medium <sup>2</sup>	DNA	Lipofectamine™ LTX Reagent	PLUS™ Reagent
96-well	$0.3 \text{ cm}^2$	100 µl	$1.0 \times 10^4$	20 µl	100 ng	0.25 - 0.45 µl	0.1 µl
48-well	$1 \text{ cm}^2$	200 µl	$2.5 \times 10^4$	40 µl	200 ng	0.5 - 0.9 µl	0.2 µl
24-well	$2 \text{ cm}^2$	500 µl	$5 \times 10^{4}$	100 µl	500 ng	1.25 - 2.25 µl	0.5 µl
12-well	$4 \text{ cm}^2$	1 ml	$1.0 \times 10^5$	200 µl	1 μg	2.5 - 4.5 µl	1.0 µl
6-well	$10 \text{ cm}^2$	2 ml	$2.5 \times 10^5$	500 µl	2.5 µg	6.25 – 11.25 µl	2.5 µl

<sup>&</sup>lt;sup>1</sup>Surface areas may vary depending on the manufacturer.

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 $<sup>^{2}</sup>$ If the volume of Lipofectamine  $^{^{\intercal}}$  LTX Reagent is too small to dispense accurately, and you cannot pool dilutions, predilute Lipofectamine  $^{^{\intercal}}$  LTX Reagent 10-fold in Opti-MEM® I Reduced Serum Medium, and dispense a 10-fold higher amount (should be at least 1.0 µl per well). Discard any unused diluted Lipofectamine  $^{^{\intercal}}$  LTX Reagent.