thermo scientific



WHITE PAPER 73104

Column properties that make an impact on Ion Chromatography

Authors

Carl Fisher and John Guajardo, Thermo Fisher Scientific, Sunnyvale, CA, USA

Keywords

Ion exchange chromatography, stationary-phase architecture, functionalization, column capacity, hydroxide-selective column

Executive summary

For the separation of ionic species, ion chromatography (IC), a type of liquid chromatography (LC), is the method of choice. The most critical component of this technique is the separation column, which is selected based on factors that include the specific analytes of interest, the sample type, and the required detection levels. This White Paper outlines the column parameters that impact the separation of charged species in solution using ion-exchange chromatography and the developments that have continued to redefine what is possible with an IC system.

Role of columns in Ion Chromatography

The characteristics of a separation column are one of the most critical considerations for obtaining optimal results from an IC system (Figure 1). The column dictates how well the ionic components of a solution are separated, which is essential for confident peak integration. It determines whether analytes will be only crudely detected or quantified in the parts per billion (ppb) range.



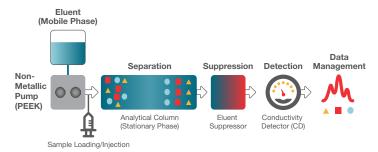


Figure 1. Schematic diagram of an IC system. Polyetheretherketone, PEEK.

The quality of the separation is influenced by multiple factors, some of which can be varied by adjusting the method parameters and others that are fixed by the properties of the column. Chromatographic conditions that the system operator has direct control over include eluent concentration, flow rate, temperature, and sample loading volume/concentration. Because IC is limited to just a few eluents, choosing a column with the right selectivity is critical for achieving a desired separation, especially when analytes are of the same charge. The initial column selection is the focus of this White Paper, and the characteristics that will be discussed are particle size, surface area, resin surface chemistry, and format. The interplay of the instrument method variables along with the selected column's stationary phase dictates how well the analytes are separated, ultimately determining the quality of the chromatographic data obtained and the quantification that can be attained.

Early IC column development

The earliest IC separations used a lightly sulfonated (R-SO₃⁻) styrene-divinylbenzene (DVB) polymer (stationary phase) to create a cation-exchange surface and dilute hydrochloric acid as the eluent (mobile phase).¹ Styrene-based polymers were preferred over the silica-based materials that were commonly used in High Performance Liquid Chromatography (HPLC) due to their superior chemical stability. The first widely used anion-exchange stationary phases were created by coating a surface-sulfonated styrene-DVB resin with a suspension of colloidal ion-exchange resin bound to the cation-exchange surface.

These columns originally used phenate-based eluents, but a hydroxide-based mobile phase was a more attractive option because of the low background achievable and the potential for use in gradient elution. However, the low eluting strength of hydroxide eluents meant high concentrations were needed to displace all the anionic analytes that were bound to the resin. This required frequent offline regeneration of the packed bed suppressors that were in use at the time, causing a bottleneck for throughput. Another concern with the use of manually prepared hydroxide-based eluents was the inability to eliminate carbonate impurity, which, by itself, has a strong analyte displacing potential, creating an unpredictable elution profile. When carbonate-based eluents were tried their strong elution potential meant that lower concentrations could be used, and, thus, fewer suppressor regenerations were required. This relative ease of use resulted in their widespread adoption and production of columns with peak separations optimized for use with carbonate eluents. As suppressor technology matured to the point where offline regeneration was no longer required and adequate suppression capacity was available, hydroxide-based eluents became a much more favorable mobile phase because of their ability to be suppressed to water. This is the ideal background for conductivity measurements, producing the best signal/ noise ratio to achieve the lowest limits of detection with conductivity detection (CD). With the development of electrolytic eluent generation (Reagent-Free IC, RFIC), on-demand production of hydroxide-based eluent from deionized water meant that carbonate contamination was no longer an issue. An added benefit is that highly reproducible gradients, which focus peaks to yield greater chromatographic peak capacity, can be produced without the need for a proportioning pump.

Achieving greater efficiency: Resin particle size

The first resins used for IC were approximately $35 \ \mu m$ in diameter. Modern IC stationary phase particles from Thermo Scientific range in size from approximately 9 to $4 \ \mu m$. The development of smaller size particles in both HPLC and IC columns was driven by the increased column efficiency that is obtained, and extension of the range of flow rates at which this is maintained, giving users more flexibility in optimizing their method. What this means to the chromatography is taller and narrower peaks that are better separated, resolving closely eluting analytes that could previously not be distinguished and facilitating peak integration, which leads to more accurate quantification.³

A consequence of developing ever smaller sized particles is the need to develop, in parallel, instrumentation that can handle the higher backpressures that arise. <u>Thermo Scientific[™] Dionex[™] High Pressure IC</u> (HPIC) systems meet this challenge with pumps, valves, and consumables that can tolerate backpressures of up to 5000 psi, which allow users to take full advantage of a column's potential. While some vendors reduce the column length to achieve shorter run times, Thermo Scientific combines a shorter column with smaller particle resins to not only achieve faster run times, but also maintain chromatographic efficiency and resolution (Figure 2).

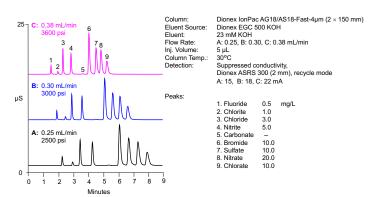
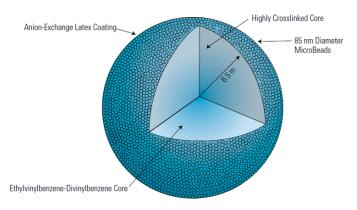


Figure 2. Reduction in run time achieved using higher flow rates in combination with a reduced length, 4 µm resin particle column.

Increasing column capacity: resin porosity

The initial polystyrene-DVB particles used for IC were nonporous and resulted in columns with relatively low capacity. To handle high analyte concentrations, improve the resolution of weakly retained ions, analyze high-ionic strength samples, and to load a higher sample volume without overloading and peak broadening, high column capacity is needed. To achieve this, resins have been engineered with varying amounts of porosity, allowing more surface area per particle to be exposed to the mobile phase and, thereby, providing more opportunities for sample analytes to bind to the surface, increasing capacity. This is exemplified by the change in core resin used for Thermo Scientific[™] Dionex[™] IonPac[™] AS11-HC particles from microporous as in Thermo Scientific™ Dionex[™] IonPac[™] AS11 (<10 Å pores, Figure 3A) to supermacroporous (2000 Å pores, Figure 3B). The larger pores present more surface for the agglomeration (attachment) of latex particles (80 nm Thermo Scientific™ Dionex[™] MicroBead[™] resin), increasing the column capacity from 45 μ eq to 290 μ eq (>500%) for a 4 \times 250 mm column.

A. Dionex IonPac AS11 resin particle



B. Dionex IonPac AS11-HC resin particle

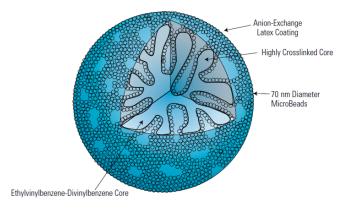
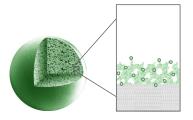


Figure 3. Comparison of Dionex IonPac AS11 and AS11-HC resin particle structures.

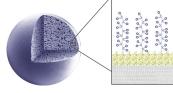
Altering selectivity: Resin surface modification

The surface of organic polymers used in anion-exchange resins can be functionalized (i.e., have their properties altered) using a two-step process of 1) addition of a chloromethyl group to the aromatic resin skeleton, followed by 2) amination with a tertiary amine. For cation-exchange resins, functionalization consists primarily of surface sulfonation via reaction with concentrated sulfuric acid or carboxylation via reaction with carboxylic acid (see reference 4 for a comprehensive review).

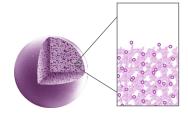
While the first stationary phases consisted of lightly sulfonated (R-SO⁻) polystyrene-divinylbenzene (PS/DVB) polymer, the surface properties of IC resins used today are modified by employing multiple strategies that include encapsulation, polymeric grafting, chemical derivatization, step-growth, and agglomeration (Figure 4). The precise way in which the stationary phase is synthesized and functionalized enables the selectivity of each resin to be finely tuned to meet specific application requirements.



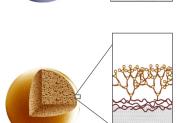
A. Polymer encapsulated substrates Examples: Metrohm Metrosep C4 Metrohm Metrosep C6



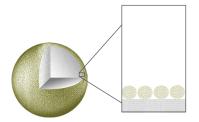
B. Polymer grafted film on porous substrates Examples: Dionex IonPac CS12A through CS20



C. Chemically derivatized polymeric substrates Examples: Shodex IC SI-52 4E Metrohm Metrosep A Supp 5 Metrohm Metrosep A Supp 7

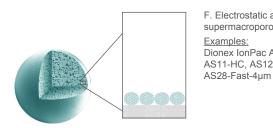


D. Step-growth polymers on polymeric substrates Examples: Dionex IonPac AS19 through AS32-Fast-4µm



E. Electrostatic agglomerated nonporous substrates Examples: Dionex IonPac AS3, AS4, AS4A, AS5, AS5A, AS7, AS9, AS11, AS17-C, CS3, CS10, CS11

Figure 4. Commonly used IC column stationary-phase architectures.



F. Electrostatic agglomerated supermacroporous substrates Examples: Dionex IonPac AS9-HC, AS10, AS11-HC, AS12A, AS16, AS18,

Column formats

Separation columns are available in multiple formats that vary by inner diameter (i.d.) and length. The column diameter used is primarily determined by the chromatographic system's properties, the sensitivity required, and the sample volume (Table 1). Most standard IC systems can run analytical (4 mm i.d.) and microbore (2 mm i.d.) columns. However, <u>Thermo Scientific[™] Dionex[™]</u> <u>ICS-4000</u> and <u>Thermo Scientific[™] Dionex[™]</u> <u>ICS-6000</u> HPIC systems are the only IC systems currently available that are capable of running capillary (0.4 mm i.d.) columns. The pumps used in both systems reliably and precisely deliver flow rates in the microliter per minute range. They are equipped with components such as suppressors, eluent generators, degas assemblies, and carbonate removal devices that are specifically designed for capillary flow rates and volumes. The low sample volumes and eluent consumption of <u>capillary IC</u> result in minimal waste disposal requirements and allow the system to be run continuously. Consequently, these systems are always ready to run samples because there is no need for extensive equilibration prior to the start of a run, which is required with standard bore systems that are regularly put into standby mode to save eluent and reduce waste. There are more than 30 Thermo Scientific capillary anion- and cation-exchange columns, many in 4 µm particle size (Table 2).

Table 1. Comparison of parameters for IC column formats.

	Analytical	Microbore	Capillary
Column i.d. (mm)	4	2	0.4
Flow Rate (mL/min)	1.0	0.25	0.01
Injection Volume (µL)	10	2.5	0.1
Eluent Consumption/Waste generated (L/month)	43.2	10.8	0.432
Best suited for	• Established procedures with specific requirements that cannot be readily changed	 Standard applications Reduced eluent consumption and waste generation Low-level determinations 	 Minimal sample/eluent consumption; and waste generation "Always-ready," minimized turnaround time Routine applications

Table 2. Comparison of IC column resin types from various vendors.

		Vendor A	Vendor B	Thermo Scientific
Particle Size	≤ 4 µm	-	1	10
	5-12 µm	14	11	40
Capillary Columns (0.4 mm i.d.)		-	-	31
	Carbonate	8	6	11
Anion Columns (Eluent Type)	Hydroxide	-	-	24
	Other	-	2	-
Cation Columns		6	4	15
Total Columns		14	12	50

A history of innovation

From the very first "Anion Separator Column" to the present day <u>Thermo Scientific[™] Dionex[™] IonPac[™] AS32-Fast-4µm</u> column, Thermo Fisher Scientific has been dedicated to continually developing new columns to meet the ever-changing needs of our customers, who are tackling new analytical challenges every day (Figures 5 and 6, reference 5).

One example of column development driven by customer needs is the recently released <u>Thermo Scientific[™] Dionex[™]</u> <u>IonPac[™] AS31</u>. One of the limitations with the analysis of haloacetic acids (HAAs) in drinking water using U.S. EPA method 557⁶ is the analysis time of approximately one hour per sample. This limits the number of samples that can be analyzed in a day, increasing the cost per sample. Customers expressed a desire for a more economical method.

To address this need, the column chemistry was modified to change the elution order of sample constituents so that the components that were directed to waste or to the mass spectrometer for detection could be grouped more efficiently. The net result was a 39% decrease in the overall run time leading to a significant reduction in cost per sample and time to result.⁷

Another example of solid-phase modifications that resulted in greater ease of use is the optimization of the surface properties of the <u>Thermo Scientific[™] Dionex[™]</u> <u>IonPac[™] CS17</u> column resin so that it does not require the use of an organic eluent modifier to resolve amines and alkanolamines.⁸ Aside from making manual eluent preparation more straightforward, a RFIC system can now be used, which further simplifies the operation, obtaining precise, reproducible results.

Carbonate-Based Columns

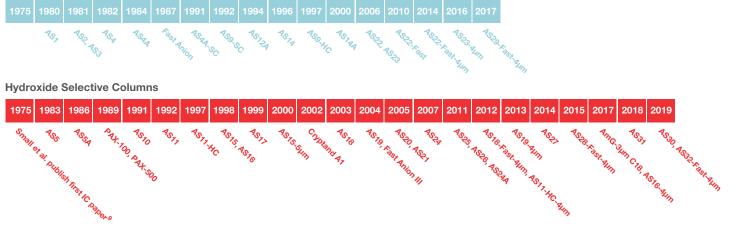
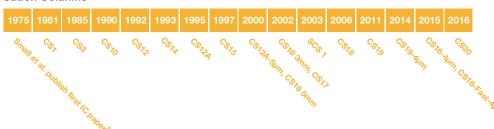
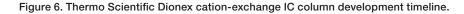


Figure 5. Thermo Scientific Dionex anion-exchange IC column development timeline.

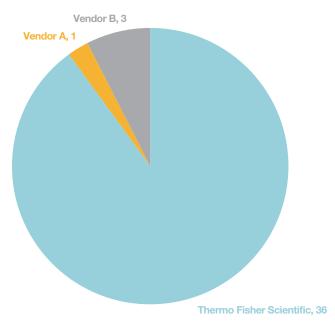


Cation Columns



thermo scientific

The history of innovation by Thermo Fisher Scientific is exemplified not only by the regularity of new column introductions each year, but also by the number of patents that they have been issued for developments in IC column technology, which is unmatched in the industry (Figure 7).



*Based on Questel Orbit Intelligence patent search tool.

Figure 7. Number of IC Column patents issued; comparison by vendor*.

Summary

The continual response to customer needs by Thermo Fisher Scientific has led to the creation of an unrivaled portfolio of IC columns that provides solutions to tackle even the most demanding chromatographic challenges.

References

- 1. Landmarks in the Evolution of Ion Chromatography, Hamish Small, Advances in Ion Chromatography, LCGC, 2013. pp. 8-15.
- 2. Recent Developments in Ion-Exchange Columns for Ion Chromatography, Chris Pohl, Advances in Ion Chromatography, LCGC, 2013. pp. 16-22.
- Thermo Scientific White Paper 70631, "Benefits of Using 4 μm Particle-Size Columns", M. Verma and C. Tanner, 2013. Available: https://assets.thermofisher.com/TFS-Assets/ CMD/Reference-Materials/wp-70631-4-micrometer-columns-wp70631-en.pdf
- 4. Weiss, J. *Handbook of Ion Chromatography;* Wiley-VCH: Weinheim, Germany, 2016; 3 vols.
- Thermo Scientific Dionex IonPac IC column selection guide, 2019. Available: https:// assets.thermofisher.com/TFS-Assets/CMD/brochures/st-70588-ic-column-selectionguide-st70588-en.pdf
- U.S. Environmental Protection Agency, Method 557, Determination of Haloacetic Acids, Bromate, and Dalapon in Drinking Water by Ion Chromatography Electrospray Ionization Tandem Mass Spectrometry (IC-ESI-MS/MS), Rev 1.0, 2009
- Thermo Scientific White Paper 72958, "Fast determination of haloacetic acids in drinking water", 2019. Available: https://assets.thermofisher.com/TFS-Assets/CMD/ Reference-Materials/wp-72958-haloacetic-acids-drinking-water-wp72958-en.pdf
- Thermo Scientific Application Update 155, "Determination of cations and amines in hydrogen peroxide by ion chromatography using a RFIC (reagent-free) system", Weerapong Worawirunwong, Sumate Peng Pumkiat, and Jeff Rohrer, 2017. Available: http://assets.thermofisher.com/TFS-Assets/CMD/Application-Notes/au-155-ic-cationsamines-hydrogen-peroxide-au72541-en.pdf
- 9. H. Small, T.S. Stevens, and W.C. Bauman. Anal. Chem. 75, 1801–1809, 1975.

Find out more at thermofisher.com/ICColumns

For Research Use Only. Not for use in diagnostic procedures. © 2019 Thermo Fisher Scientific Inc. This information is presented as an example of the capabilities of Thermo Fisher Scientific products. It is not intended to encourage use of these products in any manners that might infringe the intellectual property rights of others. Specifications, terms and pricing are subject to change. Not all products are available in all countries. Please consult your local sales representatives for details. WP73104-EN 0919C

